

# The Influence of Epigenetic Mechanisms on the Development of Metabolic Dysfunction Associated Steatotic Liver Disease: A Review

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## ABSTRACT

Metabolic dysfunction-associated steatotic liver disease (MASLD) occupies a leading place in the structure of modern hepatology. A growing body of literature identifies MASLD as a global epidemic. Accumulated data from studies in the field of hepatology support the idea that MASLD is a hepatic manifestation of a systemic metabolic disease. MASLD is a multifactorial metabolic disease associated with the presence of insulin resistance, abdominal obesity, oxidative stress, endothelial dysfunction and a systemic inflammatory response. Current scientific data demonstrate the existence of a relationship between MASLD and an increased risk of developing cardiovascular disease, regardless of traditional risk factors such as diabetes mellitus, dyslipidemia, obesity and hypertension. The pathogenesis of MASLD includes the development of hepatic steatosis with subsequent progression to metabolic dysfunction-associated steatohepatitis, fibrosis, cirrhosis, and hepatocellular carcinoma. Epigenetics, a new field of biology that studies the influence of external factors on gene activity without changing in deoxyribonucleic acid (DNA) sequences, offers a new perspective on the pathogenesis of MASLD. This review summarizes current knowledge on the epigenetic determinants of MASLD, such as DNA methylation, histone modifications, noncoding ribonucleic acids (RNAs) and N6-methyladenosine in patients with MASLD which may also contribute to the development of preventive or therapeutic strategies for MASLD-associated complications.

**Key words:** epigenetics – metabolic dysfunction-associated steatotic liver disease – metabolic dysfunction – associated steatohepatitis – DNA methylation – histone modifications – noncoding RNAs – N6-methyladenosine.

**Abbreviations:** ALT: alanine aminotransferase; AST: aspartate aminotransferase; circRNAs: circular RNAs; DNA: deoxyribonucleic acid; DNMT: DNA methyltransferase; H3K27: 27<sup>th</sup> amino acid in histone H3; H3K9: 9<sup>th</sup> amino acid in Histone H3; HCC: hepatocellular carcinoma; HDAC: histone deacetylases; LF: liver fibrosis; lncRNAs: long noncoding ribonucleic acids; MASLD: metabolic dysfunction-associated steatotic liver disease; miRNAs: micro ribonucleic acids; mRNA: messenger ribonucleic acids; MASH: metabolic dysfunction-associated steatohepatitis; ncRNAs: noncoding ribonucleic acids; RNAs: ribonucleic acids; TC: total cholesterol; TG: triglycerides.

## INTRODUCTION

Metabolic dysfunction-associated steatotic liver disease (MASLD) is a progressive spectrum of liver diseases that begins with benign fat accumulation in hepatocytes (steatosis), followed by inflammation and hepatocellular injury – metabolic dysfunction-associated steatohepatitis (MASH) and progresses to liver

fibrosis (LF) [1]. Although MASLD is typically asymptomatic, 25% of patients with MASH may progress to liver cirrhosis, and 10% may develop decompensated liver disease [2]. MASLD is currently considered the most common liver disease in developed countries [3]. Its global prevalence in the general population is variable: in the Eastern countries it is 20–30% and in Asia 5–18% [4]. Currently, liver-related mortality constitutes the third leading cause of death in MASLD individuals [5]. The development of MASLD is closely related to the characteristics of the metabolic syndrome, such as central obesity, insulin resistance, type 2 diabetes mellitus, arterial hypertension and dyslipidemia [6]. The underlying mechanisms of metabolic diseases are multifaceted and involve

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both genetic and non-genetic (epigenetic) factors. Epigenetics, first described by Conrad Hal Waddington in 1942, refers to heritable yet reversible changes in gene expression. It influences the phenotype by regulating gene transcription without altering the primary deoxyribonucleic acid (DNA) sequence. Epigenetic changes modulated by environmental influences such as diet, physical activity, or lifestyle shape gene expression by switching genes on and off [7]. Epigenetic modifications may alter the expression of genes involved in lipid metabolism, inflammation, and oxidative stress, all of which contribute to the development of MASLD [8]. Despite the undeniable role of genetic factors in the pathogenesis of MASLD, the rising prevalence of the disease cannot be explained solely by environmental and genetic influences. Epigenetics, involving reversible and heritable changes in gene expression without altering the underlying nucleotide sequence, serves as an important mechanistic link in this phenomenon. A growing body of evidence identifies epigenetics as a crucial factor contributing to the pathogenesis and progression of MASLD [9].

According to the current understanding of MASLD pathogenesis, epigenetic changes such as DNA methylation and histone modifications may activate genes that promote lipid synthesis in hepatocytes, thereby contributing to the development of hepatic steatosis. At later stages, alterations in histone proteins promote the release of inflammatory cytokines, the development of oxidative stress, and the progression to MASH [10]. A growing body of evidence also suggests that hepatic DNA methylation in patients with MASLD is a key factor in the transition from steatosis to severe LF with the development of MASH [11]. Moreover, alterations in histone proteins trigger the development of hepatocellular carcinoma (HCC) in patients with MASLD [12]. Elucidation of the epigenetic factors that predispose individuals to MASLD may lead to the development of non-invasive biomarkers for early diagnosis and may enable preventive and therapeutic strategies for high-risk populations [13].

This review discusses current epigenetic variations associated with the development and progression of MASLD and possible therapeutic strategies aimed at correcting these variations.

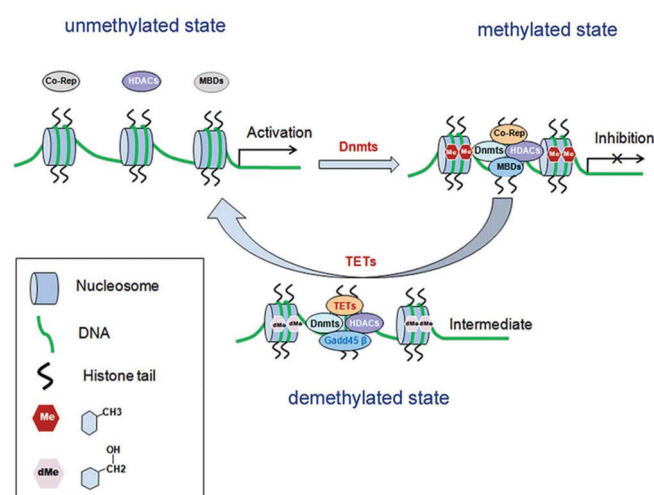
## EPIGENETIC FACTORS IN THE DEVELOPMENT OF MASLD

In 1942, the English biologist Conrad Hal Waddington proposed the new term 'epigenetics' to describe the processes by which the genotype brings the phenotype into being [14]. Epigenetics involves heritable modifications to the structure and biochemistry of chromatin, without altering the DNA sequence [15]. The epigenetic mechanisms modulate diverse physiological and pathological processes via the regulation of gene expression through alterations in epigenetic code accessibility within the chromatin [16]. Epigenetic changes can be inherited and modified by environmental factors, are considered reversible, and may offer novel personalized approaches to the prevention and treatment of a variety of diseases [17]. Cytosine and histone modifications, as well as alterations in the localization of nucleosomes occurring at the molecular level, are potential drivers of epigenetic

regulatory mechanisms [18]. Growing evidence suggests that MASLD progression is substantially influenced by multiple epigenetic mechanisms. Although some studies reported that one of the DNA methylation variants was associated with a threefold increased risk of lean MASLD compared with non-lean MASLD, the epigenetic pathogenesis of lean MASLD remains poorly understood despite extensive basic and clinical research on MASLD [19]. Epigenetic factors – including DNA methylation, histone post-translational modifications, noncoding RNAs, and N6-methyladenosine – have been associated with fatty liver disease and its severity [20].

### The Role of DNA Methylation

DNA methylation (Fig. 1) is a universal chemical modification by which methyl groups are added to the DNA molecule [21]. DNA methylation most often happens on the cytosine phosphate guanine islands, a site in which a cytosine is located next to a guanidine [22]. Mainly noted within telomeres, centromeres, repeat sequences, and inactive X-chromosomes, DNA methylation is involved in several biological processes, such as genomic imprinting, regulation of epigenetic gene expression, genome stability and transposon silencing [23].



**Fig. 1.** Schematic diagram of DNA methylation in the regulation of gene transcription. DNA methylation is mediated by DNA methyltransferases (Dnmts), which is recruited by methyl-binding domain proteins (MBDs) and form transcription repressor complexes together with co-repressors (Co-Rep) and HDACs – induces transcriptional inhibition. DNA demethylation is mediated by the ten-eleven translocases (TETs), which can catalyze oxidation of 5-methylcytosine (5mC) to 5-hydroxymethylcytosine (5hmC) leads to gene transcriptional activation. Copyright © 2015 Chen, Huang, Michael and Xiao [24]. Available via license: Creative Commons Attribution 4.0 International.

Many DNA methylations take place in the liver, so that hepatic steatosis is commonly seen from the point of view of the deregulation of carbon metabolism, being related to folate deficiency [25].

In a study of liver mitochondrial DNA methylation significantly higher methylation of mitochondrial gene nicotinamide adenine dinucleotide dehydrogenase-6 and lower

messenger RNA (mRNA) levels was reported in MASH patients in comparison to simple steatosis patients. This points towards an association of hepatic methylation and transcriptional downregulation of dinucleotide dehydrogenase-6 with the severity of MASLD [26]. In a study, using genome wide DNA methylation analysis from liver samples of 45 morbidly obese patients in different stages of MASLD, 9 MASLD associated genes were extracted exhibiting the variation in methylation [27].

Also, in a study, using epigenome-wide association studies six differentially methylated 5'-C-phosphate-G-3' sites (regions of DNA where a cytosine nucleotide is followed by a guanine nucleotide in the linear sequence of bases along its 5' → 3' direction) in genes such as *ACSL4*, *CRLS1*, *CTP1A*, *SIGIRR*, *SSBP1* and *ZNF622* were identified which could serve as serum biomarkers to distinguish between those with MASH and simple steatosis [28]. It was demonstrated that the relationship of increased hepatic dipeptidyl peptidase-4 expression levels and corresponding reduction in DNA methylation was observed in human liver biopsies to different hepatosteatosis and MASH stages which suggested a very complex intertwined regulation of fuel metabolism through epigenetic modulation [29]. In a study using epigenome-wide association studies, eight genes were identified associated with markers of liver function, among which altered DNA methylation at Solute carrier family 7 member 11 was linked with the reduced incidence of hepatic steatosis, through a favorable association with a panel of genes involved in lipid metabolism [30]. In an epigenomic analysis, which used data from the Infinium Methylation EPIC array from 325 individuals with MASLD, it was demonstrated that DNA methylation is a mechanism underlying changes in the cellular composition of hepatocytes in the pathogenesis of MASLD, including the development of LF [31].

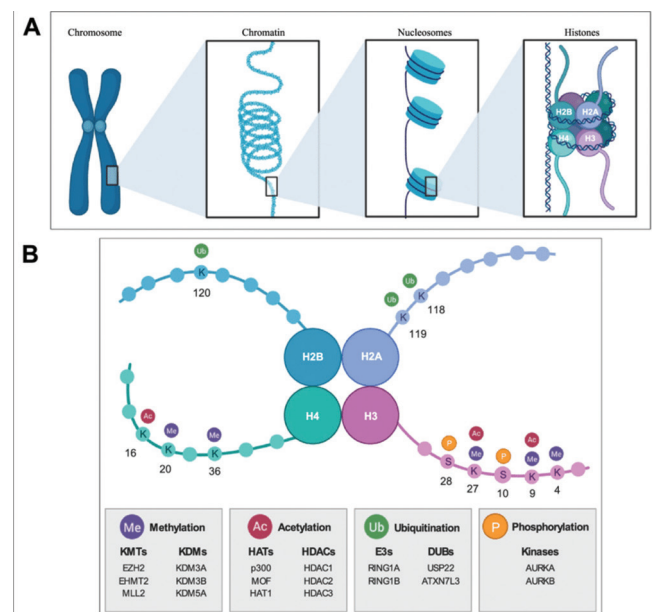
Recent studies suggest that altered gene expression due to abnormal DNA methylation may also be involved in the progression of MASLD to HCC. In a study investigating liver gene expression along with DNA methylation changes in mice using the stelic animal model of MASH derived hepatocarcinogenesis, it was found that aberrantly expressed genes had greater accrual of DNA methylation abnormalities with the majority of atypically methylated genes found in fully developed HCC. One of these genes, tubulin beta 2B class IIB, was found hypomethylated and overexpressed as liver carcinogenesis progressed [32]. Another study, using relevant mice models (diet induced MASLD mice, stelic animal model of MASH derived HCC as well as choline and folate deficient diet mice models exhibiting MASLD to HCC progression that resemble carcinogenesis in humans) characterized a progressive increase in glycine N-methyltransferase promoter methylation and a corresponding reduction of glycine N-methyltransferase expression [33]. In a meta-analysis decreased mRNA levels of *patatin-like phospholipase domain-containing protein 3* while increased expression of *PARVB* (a protein that in humans is encoded by the *PARVB* gene) was correlated with MASLD succession to HCC [34].

Thus, numerous studies in human and animal models suggest altered DNA methylation patterns at global and locus specific levels leading to hepatic lipid accumulation, steatosis, inflammation, and injury responsible for establishment and progression of MASLD [35]. The study of DNA methylation

is a promising field of modern science for understanding the influence of epigenetic factors on the development and progression of MASLD.

### The Role of Histone Modification

DNA within cells is packaged as chromatin, a dynamic structure composed of nucleosomes as the fundamental building blocks. Histones are the central component of the nucleosomal subunit, forming an octamer containing the four core histone proteins (H3, H4, H2A, H2B) [36]. As a component of octamer, histone undergoes several different post-translational modifications through different histone-modifying enzymes [37]. There are various types of histone modifications (Fig. 2), such as methylation, acetylation, lactylation, phosphorylation, dopaminylation, and ubiquitination, among others [38, 39].



**Fig. 2.** Schematic diagram of histone modification. A. Schematic representation of DNA-histone protein complexes. B. Schematic representation of a nucleosome. The main histone modifications (methylation, acetylation, ubiquitination and phosphorylation) are localized on the tails of the four main histones (H2B, H2A, H4, H3). Enzymes involved in the mentioned modifications are listed in gray boxes. © The Author(s) 2022 (Braghini M.R., Lo Re O., Romito I., et al.) [40]. Available via license: Creative Commons Attribution 4.0 International.

Dysregulated modifications of histone methylation contribute to functional abnormalities that exacerbate the progression of various diseases, including diabetes, hypertension, atherosclerosis, fatty liver disease, tumors and autoimmune disorders [41, 42]. In recent years, the role of histone methylation in MASLD has attracted increasing attention. In a study it was reported that increased trimethylation of the 27<sup>th</sup> amino acid in histone H3 (H3K27) affects the increased expression of genes involved in lipid synthesis. Enhancer of zeste 2 polycomb repressive complex 2 subunit, identified as the specific methyltransferase responsible for H3K27 methylation, plays a significant role in modulating diverse phenotypes of MASLD and operates through distinct

gene targets at different stages of the disease progression [43]. In a study the inbred strain of laboratory mouse models (C57BL/6J and DBA/2J), induced by an adipose-derived methyl-deficiency diet, manifest specific phenotypic changes characteristic of MASH. The study reported that these changes were concomitant with variations in the levels of 9<sup>th</sup> amino acid in Histone H3 (H3K9), H3K27, and 20<sup>th</sup> amino acid in Histone H4 methylation, underscoring the pivotal role of histone methylation in the initiation and advancement of MASH [44]. Schuster et al. [45] reported that histone methylation exerted an influence not only on the acute physiological alterations that underlie the transition from liver steatosis to MASH, but also directly modulated factors implicated in liver inflammation, including hepatocyte lipotoxicity, mitochondrial dysfunction, endoplasmic reticulum stress, and other related mechanisms of the MASLD development. Also in a study it was shown that histone demethylation was involved in the development of MASLD [46]. The study demonstrated that lysine demethylase 7A overexpression could erase the H3K9me2 and H3K27me2 repressive markers on the diacylglycerol O-acyltransferase 2 promoter, thereby increasing the expression of diacylglycerol O-acyltransferase 2 and triglycerides accumulation, which, finally, induced hepatic steatosis. As Stearoyl-CoA desaturase 1 and diacylglycerol O-acyltransferase 2 enzymes are potential targets for the treatment of MASLD and clinical trials are ongoing, PHD Finger Protein 2 and lysine demethylase 7A could provide potential therapeutic targets in treating MASLD [47].

A few studies further suggested that histone acetylation can be a potential target for MASLD. The active phosphorylated form of FTY720/fingolimod, a prodrug treating multiple sclerosis, could reduce fatty acid synthase expression by histone acetylation alteration, inhibit ceramide production and hepatic steatosis in diet-induced MASLD mice [48, 49]. Interestingly, nuclear receptor subfamily 2 group F member 6 expression was increased in the livers of MASLD patients and reduced by metformin treatment in obese mice. Therefore, nuclear receptor subfamily 2 group F member 6 antagonists might offer a therapeutic approach for treating MASLD through histone acetylation [50]. It has also been reported that histone acetylation can be simultaneously involved in the regulation of multiple genes. The homozygous knock-in of a serine-to-alanine mutation at Phospho-Caspase 9 in Liver X receptor alpha could induce liver steatosis but prevent cholesterol accumulation, inflammation and fibrosis, thereby slowing the development from simple hepatic steatosis to MASH [51].

Ubiquitination (the covalent attachment of ubiquitin to acceptor residues in proteins) and sumoylation (the conjugation of any small ubiquitin-like modifiers member to a substrate) are recently demonstrated to be novel forms of histone modification. Studies have reported that post-translational modifications of transcription factors during protein processing play an important role in controlling many biological events [52]. In a study investigating the hepatic gene networks in obese patients with MASLD, hepatic fibrosis signaling was found to be the most significant pathway in the up-regulated MASLD gene cluster, whereas the endoplasmic reticulum stress and protein ubiquitination pathways to be the most significant pathways in the down-regulated MASLD gene cluster [53]. Besides ubiquitination, transcription factors

can undergo several types of protein post-translational modifications, including acetylation, phosphorylation, and glycosylation. Currently, little is known about the role of these factors in the development of MASLD [54]. There are also relatively few reports on the role of histone ubiquitination and phosphorylation in the development and progression of MASLD [55].

### The Role of Noncoding RNAs

Noncoding RNAs (ncRNAs) constitute a substantial part of the transcriptome and lack discernible protein-coding functions. However, ncRNAs have been involved in a wide range of biological processes, including disease pathogenesis [56]. Advancements in sequencing technology and data analysis have allowed researchers to discover numerous ncRNAs, including long noncoding RNAs (lncRNAs) [57], circular RNAs (circRNAs) [58] and small ncRNAs [59]. One subgroup of small ncRNAs – microRNAs – are endogenous single-stranded RNAs that play a crucial role in the regulation of biological processes and epigenetic mechanisms [60]. Recently, experimental studies have appeared that link aberrant function of lncRNAs with the development and progression of MASLD. In a study alu-mediated p21 transcriptional regulator was first identified in a search for human lncRNAs involved in cell proliferation [61]. The alu-mediated p21 transcriptional regulator was later found to be significantly upregulated in the fibrotic liver that was experimentally induced by repeated exposure to carbon tetrachloride and in mice with bile duct ligation, as well as humans with LF [62]. LF-associated lncRNA-1 was first identified in a microarray analysis to profile lncRNAs in carbon tetrachloride-treated mice, with increased expression occurring in hematopoietic stem cells [63]. In a study LF-associated lncRNA-1 was also found to promote intrahepatic cholangiocarcinoma proliferation and invasion through a similar pathway [64]. Most of the lncRNAs that have been implicated in MASH have been identified using animal models of hepatic fibrosis, largely carbon tetrachloride-induced fibrosis. Carbon tetrachloride is a common method for inducing LF, and like fibrogenesis attributed to MASLD in humans, it causes hematopoietic stem cell activation, dysregulated extracellular matrix production and degradation, and progressive hepatic fibrosis [65]. Several lncRNAs have been recently described to be clinically relevant in HCC [66]. The lncRNA activating regulator of dickkopf-related protein 1 was markedly upregulated in patients with HCC and HCC-derived cell lines. Upregulation of lncRNA activating regulator of dickkopf-related protein 1 was associated with poor patient survival [67]. Xu et al. [68] investigated lncRNAs with potential functions in cell cycle by analyzing RNA-sequencing data of tumor and adjacent healthy tissue from patients with HCC. lncRNA-mRNA coexpression analysis revealed that the DDX11-AS1 (an lncRNA located on chromosome 12p11), was highly upregulated in HCC and its covarying genes were associated with cell cycle progression.

Studies have shown that circRNAs and N6-methyladenosine had a critical role in the development of various diseases including obesity and MASLD [69]. circRNA may also play a role in a non-invasive approach to predict MASLD [70]. A growing body of evidence suggested that aberrant expression

of circRNAs was linked to the occurrence and development of liver diseases, including HCC, liver regeneration, and hepatic steatosis [71-73]. Guo X.Y. conducted a major survey, reporting that circRNA-0046367 expression was downregulated during hepatocellular steatosis *in vivo* and *in vitro* and its normalization abolished the inhibitory effects exerted by microRNA 34a on peroxisome proliferator-activated receptor  $\alpha$  (PPAR $\alpha$ ) by blocking the miRNA/mRNA interaction with miRNA response elements [74]. CircRNA-002581 expression was significantly upregulated in MASH cell and mouse models. In a study was demonstrated that circRNA-002581 knockdown markedly attenuated hepatic lipid accumulation, oxidative stress, and inflammation (as evidenced by decreased tumour necrosis factor  $\alpha$  (TNF $\alpha$ ) and interleukin-6 expression) [75]. In addition, new data suggested a link between circRNA and the development of LF in patients with MASLD. It was reported that changes in the expression profile of circRNAs could promote or suppress hepatic fibrosis by regulating the activation and proliferation of hepatic stellate cells [76]. CircRNAs have become a new research hotspot in the field of MASLD in recent years. Many circRNAs with important physiological and clinical significance have been identified. Given their stability, tissue specificity, and functional activity, circRNAs are promising to as a non-invasive diagnostic tool and therapeutic target for MASLD [77].

Another subtype of ncRNAs that are involved in the fundamental pathological process that regulates the progression of simple steatosis to steatohepatitis, cirrhosis, and ultimately HCC are microRNAs (miRNAs) [78]. The miRNAs are short (19–23 nucleotides) noncoding RNA molecules that confer a crucial layer of regulation of gene expression via mRNA destabilization, translational repression, or activation of transcription/translation [79]. These molecules are receiving increasing attention since they are commonly deregulated in pathological situations, and are currently the most intensely studied epigenetic factors in MASLD [80, 81]. A study was demonstrated that altered expression of miRNAs was associated with liver metabolism dysregulation, liver injury, LF and tumour development, making miRNAs attractive therapeutic strategies for the diagnosis and treatment of liver diseases [82].

The most expressed miRNA in human liver - miR-122 - is inhibited in steatohepatitis, acting as a suppressor of tumors in the liver [83]. A study demonstrated that silencing of miR-122 was an early event during hepatocarcinogenesis from MASH, and miR-122 could be a novel molecular marker for evaluating the risk of HCC in patients with MASH [84]. Also, other miRNAs have been identified to be involved in MASLD progression. Thus, Pirola et al. [85] identified significant fold differences in serum levels of miR-192 (4.4-fold change in MASH vs. controls). Cermelli et al. [86] identified increased serum levels of miRNA-34a and miRNA-16 in MASLD patients compared to controls [86]. Yamada et al. [87] identified in a cohort of MASLD diagnosed patients increased serum abundance of miRNA-21, miRNA-34a and miRNA-451 relative to controls [87]. Tan et al. [88] identified miRNA-1290, miRNA-27b-3p and miRNA-192-5p as a panel of high diagnostic accuracy for MASLD. Guo et al. [89] reported three miRNAs (miRNA-301a-3p, miRNA-34a-5p

and miRNA-375) as potential biomarkers to assess the severity of MASLD. In addition, several studies have evaluated the contribution of miRNA-34a upregulation to the progression of LF and the development of HCC. In a study using a chemically induced rat model of LF combined with *in vitro* experiments in human hepatocytes and co-cultured hematopoietic stem cells, it was demonstrated that the miRNA-34a/SIRT1/p53 signalling pathway was activated specifically in hepatocytes and contributed to hepatocyte apoptosis and subsequent hematopoietic stem cell activation [90]. Thus, MASLD is associated with changes in hepatic miRNAs expression patterns at early, intermediate and late stages, and specific miRNAs species appear to be involved in steatosis development and steatosis progression to MASH and LF [91]. Further research is required to investigate the impact of ncRNAs on the development and progression of MASLD, as well as to explore the therapeutic potential of their use in the treatment of this cohort of patients.

### The Role of N6-methyladenosine RNA Modification

The N6-methyladenosine modification has been increasingly described to have a dense association with the pathological machinery of various diseases [92]. The N6-methyladenosine RNA modification may highly regulate hepatic function and the development of liver diseases, providing some directions to understand the mechanism of malicious activities in the development of liver diseases [93]. A study found that N6-methyladenosine modification might be closely associated with the initiation and/or development of obesity and MASLD, which might progress to end-stage liver disease [94]. Another study demonstrated that N6-methyladenosine modification might play a role in the formation of specific and complexed microenvironments in the liver [95]. Recent studies have shown that N6-methyladenosine modification might be a key player in the pathogenesis of MASLD, which may provide new mechanistic insight into this disease [96]. The roles of N6-methyladenosine RNA methylation in the pathogenesis of liver diseases were also explored, providing new insights for studying the molecular mechanism of liver diseases [97]. However, the exact mechanism and role of N6-methyladenosine-modified RNAs in development and progression of MASLD remains poorly understood.

## EMERGING THERAPEUTIC STRATEGIES TARGETING EPIGENETIC MECHANISMS

In recent years, there has been growing interest among researchers in elucidating the role of epigenetic modifications in the treatment of MASLD.

Considering the essential function of epigenetic enzymes in the regulation of inflammation associated with MASH progression, epigenetic inhibitors could hold significant promise for the exploration and development of innovative treatments for this condition [98]. In contrast to traditional therapies, drugs targeting epigenetic-modifying enzymes have been developed with a focus on gene regulation [99]. These drugs are categorized into four main types based on their mechanisms: DNA methyltransferase (DNMT) inhibitors,

histone deacetylases (HDAC) inhibitors, and miRNA inhibitors.

### DNMT Inhibitors

In the early 1960s, two nucleoside DNMT inhibitors were discovered. These were 5-azacytidine (azacitidine, AZA, Vidaza) and its derivative, 5-2'-deoxycytidine (decitabine, DAC, Dacogen). Although the majority of DNMT inhibitors therapeutic effects are focused on oncology, numerous studies now explore their role in adipocyte biology. Treatment with decitabine induces significant hepatic DNA hypomethylation in mice fed a high-fat diet, thereby leading to a marked reduction in hepatic lipid accumulation [100, 101].

Pant et al [102] demonstrated that treatment of MASLD mice with DNMT inhibitors (azacitidine and zebularine) ameliorated different metabolic biochemical parameters (decrease the levels of plasma glucose, total cholesterol (TC), serum glutamic oxaloacetic transaminase, serum glutamic pyruvic transaminase) and increased insulin sensitivity, preventing the progression and development of MASLD [102]. This finding lays the groundwork for developing epigenetic-based therapeutics for the prevention and treatment of MASLD and other metabolic disorders. In another study, treatment with the DNA methylation inhibitor 5-Aza-2'-deoxycytidine significantly reversed DNA methylation levels and improved lipid accumulation in MASLD rat models [103].

Recently, some new nucleoside DNMT inhibitors and nonnucleoside DNMT inhibitors, including hydralazine, procaine and MG98, have been identified and are currently being investigated [104]. In a study hydralazine treatment significantly attenuated the progression of LF [this led to moderate alleviation of hepatocyte degeneration, significantly decreased LF, moderately ameliorates insulin resistance, decreased the serum and hepatic triglycerides (TG) levels] in rats with hypertension and steatohepatitis. Research data suggest that hydralazine shows promise as a potential treatment for MASH exacerbated by hypertension [105].

Current evidence suggests that epigenetic therapies demonstrate promise as a treatment of the terminal stage of MASLD – HCC. In the study, DNMT inhibitors irreversibly bound to DNMT1 and partially restored hepatocytic differentiation in HCC cells, making them less tumorigenic [106]. Despite numerous studies, administration strategies for DNMT inhibitors as a treatment for MASLD still need to be thoroughly evaluated in carefully designed clinical trials.

### HDAC Inhibitors

HDAC inhibitors have received considerable attention in the field of hepatology, probably owing to their antiproliferative properties and ability to induce cell death via deacetylation of multiple HDAC substrates [107]. Increasing evidence suggests that HDAC1 and HDAC2 (members of the HDAC family) are involved in the regulation of hepatocyte death, thereby implying that HDAC1 and HDAC2 serves a key role in the pathogenesis of liver disease. In this study, the roles of HDAC inhibitors (trichostatin A, sodium butyrate, and MS-275) in mouse hepatocyte apoptosis were examined. It was shown that HDAC inhibitors suppressed hepatocyte apoptosis and increase cell viability [108]. Valproate sodium, a broad class I and II

HDAC inhibitor, has also been shown to block myofibroblast differentiation and fibrogenesis in mouse models of LF [109]. Another study found that HDAC11, a subtype of the HDAC family, was markedly overexpressed in both in vitro and in vivo models of MASLD. In this study, a novel hydrazide-based HDAC11 inhibitor was found to inhibit de novo lipogenesis and promote fatty acid oxidation, thus mitigating hepatic lipid accumulation and pathological symptoms in MASLD mice [110].

Consequently, although there is considerable interest in developing HDAC inhibitors as therapeutic agents for liver diseases, particularly MASLD, further investigation is required to assess potential side effects, design more target-selective inhibitors, and evaluate their efficacy in patients.

### miRNA Inhibitors

Over the last few decades, numerous studies have demonstrated that inhibition of miRNAs can control the development of MASLD. miRNA-34a antagonists significantly restored the transmembrane potential of cellular mitochondria, improved cellular mitochondrial function, and reduced fat accumulation in the construction of MASLD mouse models [111].

In the meta-analysis, in which miR-34a antagonists were used as an intervention, hepatic TG, TC, alanine aminotransferase (ALT), and aspartate aminotransferase (AST) levels were significantly reduced, supporting the hypothesis that endogenous miRNA-34a is detrimental to MASLD and MASH. Inhibition of miRNA-34a activated fatty acid oxidation, and inhibited hepatocyte steatosis. It was also found that inhibition of hepatic miRNA-34a reduced hepatocyte reactive oxygen species levels and decreased gene expression of fibrogenesis [112].

Using complementary animal models of MASH, the study demonstrated that miRNA-21 ablation significantly reduces liver steatosis, inflammation and fibrosis [113]. Wang et al. [114] injected miRNA-21 antagonists into MASLD mice, and the levels of serum lipids (TG, TC, low-density lipoprotein) and transaminases (ALT, AST) were decreased. The expression of lipid synthesis-related genes was inhibited, confirming that fat accumulation and inflammation associated with MASLD progression to MASH can be reduced by inhibition of miRNA-21 [114]. In further animal tests, when MASLD mice were given miRNA-103a-3p inhibitors, this significantly decreased the serum TG, TC, ALT, and AST levels of the mice [115]. The administration of miRNA-103-3p antagonists reduced lipid droplet aggregation and significantly reduced inflammatory responses, abnormal lipid metabolism, and oxidative stress in MASLD mice. Therefore, inhibition of miRNA-103-3p offers a potential therapeutic strategy for treating MASLD [116].

Despite the promising therapeutic effects of miRNA inhibitors, there is still a scarcity of evidence to support the efficacy of individual miRNAs-based treatments for MASLD and MASH. More carefully designed preclinical studies involving miRNAs-inhibitor therapies in MASLD and MASH are essential to advance miRNA-based therapies for patients with these diseases.

## CONCLUSIONS AND FUTURE PERSPECTIVES

This review summarizes the current knowledge regarding epigenetic determinants of MASLD. Genetic variation explains only a small fraction of environmental and hereditary disease risks, whereas epigenetic modifications, such as DNA methylation, histone modifications, noncoding RNAs, N<sup>6</sup>-methyladenosine, affect the majority of MASLD phenotypes. The research into the potential role of epigenetics in MASLD is still in its infancy and needs to be improved. There is currently considerable interest in identifying epigenetics-based therapeutic strategies to prevent the development of MASLD-related conditions. Despite the fact that there is still limited knowledge, current evidence suggests that epigenetic therapies also offer promise as a treatment of MASLD. However, administration strategies for incorporating epigenetic drugs into routine clinical practice still need to be thoroughly evaluated in further clinical studies.

**Conflicts of interest:** None to declare.

**Authors' contributions:** N.Z. conceived and designed the study. T.A. analyzed the data and drafted the manuscript. N.Z. and T.A. interpreted the data. N.Z. critically revised the manuscript, approved the final version to be published.

## REFERENCES

- Italian Association for the Study of the Liver (AISF). AISF position paper on nonalcoholic fatty liver disease (NAFLD): Updates and future directions. *Dig Liver Dis.* 2017;49(5):471-483. doi:10.1016/j.dld.2017.01.147
- Wong VW, Wong GL, Choi PC, et al. Disease progression of non-alcoholic fatty liver disease: a prospective study with paired liver biopsies at 3 years. *Gut.* 2010;59(7):969-974. doi:10.1136/gut.2009.205088
- Vernon G, Baranova A, Younossi ZM. Systematic review: The epidemiology and natural history of non-alcoholic fatty liver disease and non-alcoholic steatohepatitis in adults. *Aliment. Pharmacol. Ther.* 2011;34:274-285. doi:10.1111/j.1365-2036.2011.04724.x
- Non-alcoholic Fatty Liver Disease Study Group. Lonardo A, Bellentani S, Argo CK, et al. Epidemiological modifiers of non-alcoholic fatty liver disease: Focus on high-risk groups. *Dig. Liver Dis.* 2015;47:997-1006. doi:10.1016/j.dld.2015.08.004
- Söderberg C, Stål P, Askling J, Glaumann H, Lindberg G, Marmur J, Hultcrantz R. Decreased survival of subjects with elevated liver function tests during a 28-year follow-up. *Hepatology.* 2010;51:595-602. doi:10.1002/hep.23314
- Cordero P, Campion J, Milagro FI, Martinez JA. Transcriptomic and epigenetic changes in early liver steatosis associated to obesity: Effect of dietary methyl donor supplementation. *Mol. Genet. Metab.* 2013;110:388-395. doi:10.1016/j.ymgme.2013.08.022
- Sodum N, Kumar G, Bojja SL, Kumar N, Rao CM. Epigenetics in NAFLD/NASH: Targets and therapy. *Pharmacol Res.* 2021;167:105484. doi:10.1016/j.phrs.2021.105484
- Portela A, Esteller M. Epigenetic modifications and human disease. *Nat Biotechnol.* 2010;28(10):1057-68. doi:10.1038/nbt.1685
- Jonas W, Schürmann A. Genetic and epigenetic factors determining NAFLD risk. *Mol Metab.* 2021 Aug;50:101111. doi:10.1016/j.molmet.2020.101111
- Li Y, Reddy MA, Miao F, Shanmugam N, Yee JK, Hawkins D, Ren B, Natarajan R. Role of the histone H3 lysine 4 methyltransferase, SET7/9, in the regulation of NF-kappaB-dependent inflammatory genes. Relevance to diabetes and inflammation. *J Biol Chem.* 2008 Sep 26;283(39):26771-81. doi:10.1074/jbc.M802800200
- Murphy SK, Yang H, Moylan CA, Pang H, Dellinger A, Abdelmalek MF, Garrett ME, Ashley-Koch A, Suzuki A, Tillmann HL, Hauser MA, Diehl AM. Relationship between methylome and transcriptome in patients with nonalcoholic fatty liver disease. *Gastroenterology.* 2013;145(5):1076-87. doi:10.1053/j.gastro.2013.07.047
- Mazzoccoli G, Vinciguerra M, Oben J, Tarquini R, De Cosmo S. Non-alcoholic fatty liver disease: the role of nuclear receptors and circadian rhythmicity. *Liver Int.* 2014;34(8):1133-52. doi:10.1111/liv.12534
- Li YY. Genetic and epigenetic variants influencing the development of nonalcoholic fatty liver disease. *World J Gastroenterol.* 2012;18(45):6546-6551. doi:10.3748/wjg.v18.i45.6546
- Waddington CH. The epigenotype. *Int J. Epidemiol.* 1942;41:10-13. doi:10.1093/ije/dyr184
- Shi Y, Zhang H, Huang S, Yin L, Wang F, Luo P, Huang H. Epigenetic regulation in cardiovascular disease: Mechanisms and advances in clinical trials. *Signal Transduct. Target. Ther.* 2022;7:200. doi:10.1038/s41392-022-01055-2
- Baylin SB, Ohm JE. Epigenetic gene silencing in cancer—A mechanism for early oncogenic pathway addiction? *Nat. Rev. Cancer.* 2006;6:107-116. doi:10.1038/nrc1799
- Jonas W, Schürmann A. Genetic and epigenetic factors determining NAFLD risk. *Mol Metab.* 2021;50:101111. doi:10.1016/j.molmet.2020.101111
- Portela A, Esteller M. Epigenetic modifications and human disease. *Nat. Biotechnol.* 2010;28:1057-1068. doi:10.1038/nbt.168.5
- Xu R, Pan J, Zhou W, Ji G, Dang Y. Recent advances in lean NAFLD. *Biomed. Pharmacother.* 2022;153:113331. doi:10.1016/j.biopha.2022.113331
- Pirola CJ, Sookoian S. Epigenetics factors in nonalcoholic fatty liver disease. *Expert Rev Gastroenterol Hepatol.* 2022;16(6):521-536. doi:10.1080/17474124.2020.1765772
- Wu YL, Lin ZJ, Li CC, et al. Epigenetic regulation in metabolic diseases: mechanisms and advances in clinical study. *Signal Transduct Target Ther.* 2023;8(1):98. doi:10.1038/s41392-023-01333-7
- Ma X, Kang S. Functional Implications of DNA Methylation in Adipose Biology. *Diabetes.* 2019;68:871-878. doi:10.2337/dbi18-0057
- Nishiyama A, Nakanishi M. Navigating the DNA methylation landscape of cancer. *Trends Genet.* 2021;37:1012-1027. doi:10.1016/j.tig.2021.05.002
- Chen XS, Huang N, Michael N, Xiao L. Advancements in the Underlying Pathogenesis of Schizophrenia: Implications of DNA Methylation in Glial Cells. *Front Cell Neurosci.* 2015;9:451. doi:10.3389/fncel.2015.00451
- Barnett MPG, Bermingham EN, Young W, Bassett SA, Hesketh JE, Maciel-Dominguez A, McNabb WC, Roy NC. Low folate and selenium in the mouse maternal diet alters liver gene expression patterns in the offspring after weaning. *Nutrients.* 2015;7:3370-3386. doi:10.3390/nu7053370
- Pirola CJ, Gianotti TF, Burgueño AL, Rey-Funes M, Loidl CF, Mallardi P, et al. Epigenetic modification of liver mitochondrial DNA is associated

- with histological severity of nonalcoholic fatty liver disease. *Gut*. 2013;62(9):1356–63. doi:10.1136/gutjnl-2012-302962
27. Ahrens M, Ammerpohl O, von Schönfels W, Kolarova J, Bens S, Itzel T, et al. DNA methylation analysis in nonalcoholic fatty liver disease suggests distinct disease-specific and remodeling signatures after bariatric surgery. *Cell Metab*. 2013;18(2):296–302. doi:10.1016/j.cmet.2013.07.004
  28. Wu J, Xu X, Zheng L, Mo L, Hua X, Chen Y, et al. Altered DNA methylation sites in peripheral blood leukocytes from patients with simple steatosis and nonalcoholic steatohepatitis (NASH). *Med Sci Monit*. 2018;24:6946–67. doi:10.12659/MSM.909747
  29. Baumeier C, Kaiser D, Heeren J, Scheja L, John C, Weise C, et al. Hepatic DPP4 DNA methylation associates with fatty liver. *Diabetes*. 2017;66(1):25–35. doi:10.2337/db15-1716
  30. Nano J, Ghanbari M, Wang W, de Vries PS, Dhana K, Muka T, et al. Epigenome-wide association study identifies methylation sites associated with liver enzymes and hepatic steatosis. *Gastroenterology*. 2017;153(4):1096–1106.e2. doi:10.1053/j.gastro.2017.06.003
  31. Johnson ND, Wu X, Still CD, Chu X, Petrick AT, Gerhard GS, et al. Differential DNA methylation and changing cell-type proportions as fibrotic stage progresses in NAFLD. *Clin Epigenetics*. 2021;13(1):152. doi:10.1186/s13148-021-01129-y
  32. Dreval K, Tryndyak V, de Conti A, Beland FA, Pogribny IP. Gene expression and DNA methylation alterations during non-alcoholic steatohepatitis-associated liver carcinogenesis. *Front Genet*. 2019;10:486. doi:10.3389/fgene.2019.00486
  33. Borowa-Mazgaj B, Krajka-Kuźniak V, Paluszczak J, Baer-Dubowska W. Gene expression and DNA methylation alterations in the glycine N-methyltransferase gene in diet-induced nonalcoholic fatty liver disease-associated carcinogenesis. *Toxicol Sci*. 2019;170(2):273–82. doi:10.1093/toxsci/kfz110
  34. Ryaboshapkina M, Hammar M. Human hepatic gene expression signature of non-alcoholic fatty liver disease progression, a meta-analysis. *Sci Rep*. 2017;7(1) doi:10.1038/s41598-017-10930-w
  35. Vachher M, Bansal S, Kumar B, Yadav S, Burman A. Deciphering the role of aberrant DNA methylation in NAFLD and NASH. *Heliyon*. 2022;8(10). doi:10.1016/j.heliyon.2022.e11119
  36. Audia JE, Campbell RM. Histone modifications and cancer. *Cold Spring Harb Perspect Biol*. 2016;8(4):a019521. doi:10.1101/cshperspect.a019521
  37. Park J, Lee K, Kim K, Yi SJ. The role of histone modifications: from neurodevelopment to neurodiseases. *Signal Transduct Target Ther*. 2022;7:217 doi:10.1038/s41392-022-01078-9
  38. Langan TA. Histone phosphorylation: stimulation by adenosine 3',5'-monophosphate. *Science*. 1968;162:579–80. doi:10.1126/science.162.3853.579
  39. Lepack AE, Werner CT, Stewart AF, Fulton SL, Zhong P, Farrelly LA, et al. Dopaminylation of histone H3 in ventral tegmental area regulates cocaine seeking. *Science*. 2020;368:197–201. doi:10.1126/science.aaw8806
  40. Braghini MR, Lo Re O, Romito I, et al. Epigenetic remodelling in human hepatocellular carcinoma. *J Exp Clin Cancer Res*. 2022;41(1):107. doi:10.1186/s13046-022-02297-2
  41. Yi X, Zhu QX, Wu XL, Tan TT, Jiang XJ. Histone methylation and oxidative stress in cardiovascular diseases. *Oxid Med Cell Longev*. 2022;2022:6023710. doi:10.1155/2022/6023710
  42. Cao YC, Shan SK, Guo B, Li CC, Li FX, Zheng MH, et al. Histone lysine methylation modification and its role in vascular calcification. *Front Endocrinol (Lausanne)*. 2022;13:863708. doi:10.3389/fendo.2022.863708
  43. Vella S, Gnani D, Crudele A, Ceccarelli S, De Stefanis C, Gaspari S, et al. EZH2 down-regulation exacerbates lipid accumulation and inflammation in in vitro and in vivo NAFLD. *Int J Mol Sci*. 2013;14:24154–68. doi:10.3390/ijms141224154
  44. Pogribny IP, Tryndyak VP, Bagnyukova TV, Melnyk S, Montgomery B, Ross SA, et al. Hepatic epigenetic phenotype predetermines individual susceptibility to hepatic steatosis in mice fed a lipogenic methyl-deficient diet. *J Hepatol*. 2009;51:176–86. doi:10.1016/j.jhep.2009.03.021
  45. Schuster S, Cabrera D, Arrese M, Feldstein AE. Triggering and resolution of inflammation in NASH. *Nat Rev Gastroenterol Hepatol*. 2018;15:349–64. doi:10.1038/s41575-018-0009-6
  46. Bricambert J, Alves-Guerra MC, Esteves P, Prip-Buus C, Bertrand-Michel J, Guillou H, et al. The histone demethylase Phf2 acts as a molecular checkpoint to prevent NAFLD progression during obesity. *Nat Commun*. 2018;9:2092. doi:10.1038/s41467-018-04361-y
  47. Kim JH, Nagappan A, Jung DY, Suh N, Jung MH. Histone demethylase KDM7A contributes to the development of hepatic steatosis by targeting diacylglycerol acyltransferase 2. *Int J Mol Sci*. 2021;22:11085. doi:10.3390/ijms222011085
  48. Rohrbach TD, Asgharpour A, Maczys MA, Montefusco D, Cowart LA, Bedossa P, et al. FTY720/fingolimod decreases hepatic steatosis and expression of fatty acid synthase in diet-induced nonalcoholic fatty liver disease in mice. *J Lipid Res*. 2019;60:1311–22. doi:10.1194/jlr.M093799
  49. Chung MY, Song JH, Lee J, Shin EJ, Park JH, Lee SH, et al. Tannic acid, a novel histone acetyltransferase inhibitor, prevents non-alcoholic fatty liver disease in in vivo and in vitro models. *Mol Metab*. 2019;19:34–48. doi:10.1016/j.molmet.2018.11.001
  50. Zhou B, Jia L, Zhang Z, Xiang L, Yuan Y, Zheng P, et al. The nuclear orphan receptor NR2F6 promotes hepatic steatosis through upregulation of fatty acid transporter CD36. *Adv Sci*. 2020;7:2002273. doi:10.1002/advs.202002273
  51. Becares N, Gage MC, Voisin M, Shrestha E, Martin-Gutierrez L, Liang N, et al. Impaired LXRA phosphorylation attenuates progression of fatty liver disease. *Cell Rep*. 2019;26:984–95.e6. doi:10.1016/j.celrep.2018.12.094
  52. Kim MY, Bae JS, Kim TH, Park JM, Ahn YH. Role of transcription factor modifications in the pathogenesis of insulin resistance. *Exp Diabetes Res*. 2012;2012:716425. doi:10.1155/2012/716425
  53. Gawrieh S, Baye TM, Carless M, Wallace J, Komorowski R, Kleiner DE, et al. Hepatic gene networks in morbidly obese patients with nonalcoholic fatty liver disease. *Obes Surg*. 2010;20:1698–709. doi:10.1007/s11695-010-0171-6
  54. Kim MY, Bae JS, Kim TH, Park JM, Ahn YH. Role of transcription factor modifications in the pathogenesis of insulin resistance. *Exp Diabetes Res*. 2012;2012:716425. doi:10.1155/2012/716425
  55. Shi Y, Qi W. Histone modifications in NAFLD: mechanisms and potential therapy. *Int J Mol Sci*. 2023;24(19):14653. doi:10.3390/ijms241914653
  56. Sun YM, Chen YQ. Principles and innovative technologies for decrypting noncoding RNAs: from discovery and functional prediction to clinical application. *J Hematol Oncol*. 2020;13:109. doi:10.1186/s13045-020-00945-8
  57. Quinn JJ, Chang HY. Unique features of long non-coding RNA biogenesis and function. *Nat Rev Genet*. 2016;17:47–62. doi:10.1038/nrg.2015.10

58. Memczak S, Jens M, Elefsinioti A, Torti F, Krueger J, Rybak A, et al. Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature*. 2013;495:333–8. doi:10.1038/nature11928
59. Lambert M, Benmoussa A, Provost P. Small non-coding RNAs derived from eukaryotic ribosomal RNA. *Noncoding RNA*. 2019;5:16. doi:10.3390/ncrna5010016
60. Vilimova M, Contrant M, Randrianjafy R, Dumas P, Elbasani E, Ojala PM, et al. Cis regulation within a cluster of viral microRNAs. *Nucleic Acids Res*. 2021;49:10018–33. doi:10.1093/nar/gkab731
61. Negishi M, Wongpalee SP, Sarkar S, Park J, Lee KY, et al. A new lncRNA, APTR, associates with and represses the CDKN1A/p21 promoter by recruiting polycomb proteins. *PLoS One*. 2014;9:e95216. doi:10.1371/journal.pone.0095216
62. Yu F, Zheng J, Mao Y, Dong P, Li G, et al. Long non-coding RNA APTR promotes the activation of hepatic stellate cells and the progression of liver fibrosis. *Biochem Biophys Res Commun*. 2015;463:679–85. doi:10.1016/j.bbrc.2015.06.162
63. Zhang K, Han X, Zhang Z, Zheng L, Hu Z, et al. The liver-enriched lnc-LFAR1 promotes liver fibrosis by activating TGF $\beta$  and Notch pathways. *Nat Commun*. 2017;8:144. doi:10.1038/s41467-017-00156-z
64. Chen C, Li H, Wang X, Wang L, Zeng Q. Lnc-LFAR1 affects intrahepatic cholangiocarcinoma proliferation, invasion, and EMT by regulating the TGF $\beta$ /Smad signaling pathway. *Int J Clin Exp Pathol*. 2019;12:2455–61. <http://www.ijcep.com/files/ijcep0092173.pdf>
65. Delire B, Starkel P, Leclercq I. Animal models for fibrotic liver diseases: what we have, what we need, and what is under development. *J Clin Transl Hepatol*. 2015;3:53–66. doi:10.14218/JCTH.2015.00009
66. Sommerauer C, Kutter C. Noncoding RNAs and RNA-binding proteins: emerging governors of liver physiology and metabolic diseases. *Am J Physiol Cell Physiol*. 2022;323(4). doi:10.1152/ajpcell.00232.2022
67. Jia G, Wang Y, Lin C, Lai S, Dai H, Wang Z, et al. LNCAROD enhances hepatocellular carcinoma malignancy by activating glycolysis through induction of pyruvate kinase isoform PKM2. *J Exp Clin Cancer Res*. 2021;40:299. doi:10.1186/s13046-021-02090-7
68. Xu M, Zhao X, Zhao S, Yang Z, Yuan W, Han H, et al. Landscape analysis of lncRNAs shows that DDX11-AS1 promotes cell-cycle progression in liver cancer through the PARP1/p53 axis. *Cancer Lett*. 2021;520:282–94. doi:10.1016/j.canlet.2021.08.001
69. Nakashima M, Suga N, Ikeda Y, Yoshikawa S, Matsuda S. Circular RNAs, noncoding RNAs, and N6-methyladenosine involved in the development of MAFLD. *Noncoding RNA*. 2024;10(1):11. doi:10.3390/ncrna10010011
70. Zeng Q, Liu CH, Wu D, Jiang W, Zhang N, Tang H. LncRNA and circRNA in patients with non-alcoholic fatty liver disease: a systematic review. *Biomolecules*. 2023;13:560. doi:10.3390/biom13030560
71. Yu J, Xu QG, Wang ZG, et al. Circular RNA cSMARCA5 inhibits growth and metastasis in hepatocellular carcinoma. *J Hepatol*. 2018;68(6):1214–27. doi:10.1016/j.jhep.2018.01.012
72. Li L, Guo J, Chen Y, et al. Comprehensive circRNA expression profile and selection of key circRNAs during priming phase of rat liver regeneration. *BMC Genomics*. 2017;18(1):80. doi:10.1186/s12864-016-3476-6
73. Guo XY, He CX, Wang YQ, et al. Circular RNA profiling and bioinformatic modeling identify its regulatory role in hepatic steatosis. *Biomed Res Int*. 2017;2017:5936171. doi:10.1155/2017/5936171
74. Guo XY, Chen JN, Sun F, et al. circRNA\_0046367 prevents hepatotoxicity of lipid peroxidation: an inhibitory role against hepatic steatosis. *Oxid Med Cell Longev*. 2017;2017:3960197. doi:10.1155/2017/3960197
75. Jin X, Gao J, Zheng R, et al. Antagonizing circRNA\_002581-miR-122-CPEB1 axis alleviates NASH through restoring PTEN-AMPK-mTOR pathway regulated autophagy. *Cell Death Dis*. 2020;11(2):123. doi:10.1038/s41419-020-2293-7
76. Liu W, Feng R, Li X, et al. TGF- $\beta$ - and lipopolysaccharide-induced upregulation of circular RNA PWWP2A promotes hepatic fibrosis via sponging miR-203 and miR-223. *Aging*. 2019;11(21):9569–9580. doi:10.18632/aging.102405
77. Zeng Q, Liu CH, Ampuero J, Wu D, Jiang W, Zhou L, Li H, Bai L, Romero-Gómez M, Tang H. Circular RNAs in non-alcoholic fatty liver disease: Functions and clinical significance. *RNA Biol*. 2024;21(1):1–15. doi:10.1080/15476286.2023.2290769
78. Bartel D.P. MicroRNAs: Target recognition and regulatory functions. *Cell*. 2009;136:215–233. doi:10.1016/j.cell.2009.01.002
79. Selbach M, Schwanhäusser B, Thierfelder N, Fang Z, Khanin R, Rajewsky N. Widespread changes in protein synthesis induced by microRNAs. *Nature*. 2008;455:58–63. doi:10.1038/nature07228
80. Ferreira DM, Simão AL, Rodrigues CM, Castro RE. Revisiting the metabolic syndrome and paving the way for microRNAs in non-alcoholic fatty liver disease. *FEBS J*. 2014;281:2503–24. doi:10.1111/febs.12893
81. Castro RE, Ferreira DM, Afonso MB, Borralho PM, Machado MV, Cortez-Pinto H, et al. miR34a/SIRT1/p53 is suppressed by ursodeoxycholic acid in the rat liver and activated by disease severity in human non-alcoholic fatty liver disease. *J Hepatol*. 2013;58:119–25. doi:10.1016/j.jhep.2012.08.024
82. Wang X, He Y, Mackowiak B, Gao B. MicroRNAs as regulators, biomarkers and therapeutic targets in liver diseases. *Gut*. 2021;70(4):784–95. doi:10.1136/gutjnl-2020-322526
83. Cheung O, Puri P, Eicken C, Contos MJ, Mirshahi F, Maher JW, et al. Nonalcoholic steatohepatitis is associated with altered hepatic microRNA expression. *Hepatology*. 2008;48:1810–20. doi:10.1002/hep.22569
84. Takaki Y, Saito Y, Takasugi A, Toshimitsu K, Yamada S, Muramatsu T, et al. Silencing of microRNA122 is an early event during hepatocarcinogenesis from non-alcoholic steatohepatitis. *Cancer Sci*. 2014;105:1254–60. doi:10.1111/cas.12498
85. Pirola CJ, Fernández Gianotti T, Castaño GO, Mallardi P, San Martino J, Mora Gonzalez Lopez Ledesma M, et al. Circulating microRNA signature in non-alcoholic fatty liver disease: from serum non-coding RNAs to liver histology and disease pathogenesis. *Gut*. 2015;64:800–12. doi:10.1136/gutjnl-2014-306996
86. Cermelli S, Ruggieri A, Marrero JA, Ioannou GN, Beretta L. Circulating microRNAs in patients with chronic hepatitis C and non-alcoholic fatty liver disease. *PLoS One*. 2011;6:e23937. doi:10.1371/journal.pone.0023937
87. Yamada H, Suzuki K, Ichino N, Ando Y, Sawada A, Osakabe K, et al. Associations between circulating microRNAs (miR21, miR34a, miR122 and miR451) and non-alcoholic fatty liver. *Clin Chim Acta*. 2013;424:99–103. doi:10.1016/j.cca.2013.05.021
88. Tan Y, Ge G, Pan T, Wen D, Gan J. A pilot study of serum microRNAs panel as potential biomarkers for diagnosis of nonalcoholic fatty liver disease. *PLoS One*. 2014;9:e105192. doi:10.1371/journal.pone.0105192
89. Guo Y, Xiong Y, Sheng Q, Zhao S, Wattacheril J, Flynn CR. A microRNA expression signature for human NAFLD progression. *J Gastroenterol*. 2016;51:1022–30. doi:10.1007/s00535-016-1178-0
90. Tian XF, Ji FJ, Zang HL, Cao H. Activation of the miR34a/SIRT1/p53 signaling pathway contributes to the progress of liver fibrosis

- via inducing apoptosis in hepatocytes but not in HSCs. *PLoS One*. 2016;11(7):e0158657. doi:10.1371/journal.pone.0158657
91. Hochreuter MY, Dall M, Treebak JT, Barrès R. MicroRNAs in non-alcoholic fatty liver disease: progress and perspectives. *Mol Metab*. 2022;65:101581. doi:10.1016/j.molmet.2022.101581
  92. Feng H, Yuan X, Wu S, Yuan Y, Cui L, Lin D, et al. Effects of writers, erasers and readers within miRNA-related m6A modification in cancers. *Cell Prolif*. 2023;56:e13340. doi:10.1111/cpr.13340
  93. Xu K, Sun Y, Sheng B, Zheng Y, Wu X, Xu K. Role of identified RNA N6-methyladenosine methylation in liver. *Anal Biochem*. 2019;578:45–50. doi:10.1016/j.ab.2019.05.005
  94. Petri BJ, Cave MC, Klinge CM. Changes in m6A in steatotic liver disease. *Genes*. 2023;14:1653. doi:10.3390/genes14081653
  95. Zhang N, Tian X, Yan T, Wang H, Zhang D, Lin C, et al. Insights into the role of nucleotide methylation in metabolic-associated fatty liver disease. *Front Immunol*. 2023;14:1148722. doi:10.3389/fimmu.2023.1148722
  96. Hu Y, Feng Y, Zhang L, Jia Y, Cai D, Qian SB, et al. GR-mediated FTO transactivation induces lipid accumulation in hepatocytes via demethylation of m6A on lipogenic mRNAs. *RNA Biol*. 2020;17:930–42. doi:10.1080/15476286.2020.1736868
  97. Yang L, Tian S, Zheng X, Zhang M, Zhou X, Shang Y, et al. N6-methyladenosine RNA methylation in liver diseases: from mechanism to treatment. *J Gastroenterol*. 2023;58:718–33. doi:10.1007/s00535-023-02008-4
  98. Zaiou M, Joubert O. Racial and ethnic disparities in NAFLD: harnessing epigenetic and gut microbiota pathways for targeted therapeutic approaches. *Biomolecules*. 2025;15(5):669. doi:10.3390/biom15050669
  99. Dai W, Qiao X, Fang Y, Guo R, Bai P, Liu S, et al. Epigenetics-targeted drugs: current paradigms and future challenges. *Signal Transduct Target Ther*. 2024;9:332. doi:10.1038/s41392-024-02039-0
  100. Flores-Sierra JJ, Muciño-Arellano MDR, Romo-Morales GDC, Sánchez-Palafox JE, Correa-Navarro VA, Colín-Castelán D, et al. The DNA methyltransferase inhibitor decitabine blunts the response to a high-animal fat and protein diet in mice. *J Lipid Res*. 2024;65:100586. doi:10.1016/j.jlcr.2024.100586
  101. Hasani S, Javeri A, Asadi A, Fakhr Taha M. Cardiac differentiation of adipose tissue-derived stem cells is driven by BMP4 and bFGF but counteracted by 5-azacytidine and valproic acid. *Cell J*. 2020;22:273–82. doi:10.22074/cellj.2020.6582
  102. Pant R, Kabeer SW, Sharma S, Kumar V, Patra D, Pal D, et al. Pharmacological inhibition of DNMT1 restores macrophage autophagy and M2 polarization in Western diet-induced nonalcoholic fatty liver disease. *J Biol Chem*. 2023;299(6):104779. doi:10.1016/j.jbc.2023.104779
  103. Li YY, Tang D, Du YL, Cao CY, Nie YQ, Cao J, et al. Fatty liver mediated by peroxisome proliferator-activated receptor- $\alpha$  DNA methylation can be reversed by a methylation inhibitor and curcumin. *J Dig Dis*. 2018;19(7):421–30. doi:10.1111/1751-2980.12610
  104. Hu C, Liu X, Zeng Y, Liu J, Wu F. DNA methyltransferase inhibitors combination therapy for the treatment of solid tumor: mechanism and clinical application. *Clin Epigenetics*. 2021;13:166. doi:10.1186/s13148-021-01154-x
  105. Yuan Y, Naito H, Kitamori K, Hashimoto S, Asano T, Nakajima T. The antihypertensive agent hydralazine reduced extracellular matrix synthesis and liver fibrosis in nonalcoholic steatohepatitis exacerbated by hypertension. *PLoS One*. 2020;15(12):e0243846. doi:10.1371/journal.pone.0243846
  106. Fernández-Barrena MG, Arechederra M, Colyn L, Berasain C, Avila MA. Epigenetics in hepatocellular carcinoma development and therapy: the tip of the iceberg. *JHEP Rep*. 2020;2(6):100167. doi:10.1016/j.jhepr.2020.100167
  107. Dokmanovic M, Clarke C, Marks PA. Histone deacetylase inhibitors: overview and perspectives. *Mol Cancer Res*. 2007;5:981–9. doi:10.1158/1541-7786.MCR-07-0324
  108. Lei WW, Zhang KH, Pan XC, Wang DM, Hu Y, Yang YN, et al. Histone deacetylase 1 and 2 differentially regulate apoptosis by opposing effects on extracellular signal-regulated kinase 1/2. *Cell Death Dis*. 2010;1:e44. doi:10.1038/cddis.2010.21
  109. Mannaerts I, Nuytten NR, Rogiers V, Vanderkerken K, van Grunsven LA, Geerts A. Chronic administration of valproic acid inhibits activation of mouse hepatic stellate cells in vitro and in vivo. *Hepatology*. 2010;51:603–14. doi:10.1002/hep.23334
  110. Zhang F, Yue K, Sun S, Lu S, Jia G, Zha Y, et al. Targeting histone deacetylase 11 with a highly selective inhibitor for the treatment of MASLD. *Adv Sci (Weinh)*. 2025;12(15):e202412903. doi:10.1002/adv.202412903
  111. Wen F, An C, Wu X, Yang Y, Xu J, Liu Y, et al. MiR34a regulates mitochondrial content and fat ectopic deposition induced by resistin through the AMPK/PPAR $\alpha$  pathway in HepG2 cells. *Int J Biochem Cell Biol*. 2018;94:133–45. doi:10.1016/j.biocel.2017.11.008
  112. Zhu Y, Tan JK, Wong SK, Goon JA. Therapeutic effects of microRNAs on nonalcoholic fatty liver disease (NAFLD) and nonalcoholic steatohepatitis (NASH): a systematic review and meta-analysis. *Int J Mol Sci*. 2023;24(11):9168. doi:10.3390/ijms24119168
  113. Rodrigues PM, Afonso MB, Simão AL, Carvalho CC, Trindade A, Duarte A, et al. miR21 ablation and obeticholic acid ameliorate nonalcoholic steatohepatitis in mice. *Cell Death Dis*. 2017;8(4):e2748. doi:10.1038/cddis.2017.172
  114. Wang XM, Wang XY, Huang YM, Chen X, Lü MH, Shi L, et al. Role and mechanisms of action of microRNA21 as regards the regulation of the WNT/ $\beta$ catenin signaling pathway in the pathogenesis of nonalcoholic fatty liver disease. *Int J Mol Med*. 2019;44(6):2201–12. doi:10.3892/ijmm.2019.4375
  115. Zhang M, Tang Y, Tang E, Lu W. MicroRNA103 represses hepatic de novo lipogenesis and alleviates NAFLD via targeting FASN and SCD1. *Biochem Biophys Res Commun*. 2020;524(3):716–22. doi:10.1016/j.bbrc.2020.01.143
  116. Ding J, Xia C, Cen P, Li S, Yu L, Zhu J, et al. MiR1033p promotes hepatic steatosis to aggravate nonalcoholic fatty liver disease by targeting ACOX1. *Mol Biol Rep*. 2022;49(8):7297–305. doi:10.1007/s11033-022-07515-w